Research article

PTPRC (CD45) is not associated with multiple sclerosis in a large cohort of German patients

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Abstract

Background: Since contradictory results have been reported, we reanalysed the 77C \rightarrow G transition in exon 4 of the protein-tyrosine phosphatase receptor-type C (*PTPRC* also known as CD45) in a large cohort of German MS patients and controls. Different isoforms of the protein are expressed, depending on alternative splicing of exons 4 (CD45RA), 5 (CD45RB) and 6 (CD45RC) (CD45RO, exons 4–6 spliced out). The 77C \rightarrow G transition does not change the amino acid sequence, but it is probably part of a motif necessary for splicing leading to the isoform CD45RA. The expression of CD45RA is increased in 77C/G heterozygous individuals. The aim of the study was to clarify the importance of the *PTPRC* 77C \rightarrow G transition in our German cohort of MS patients.

Methods: PCR products of exon 4 were digested using endonuclease *Mspl*. The resulting restriction fragments of the wildtype C allele are 198 and 62 bp in length. In the G allele an additional restriction site is present yielding fragments of 114 and 84 bp.

Results: The G allele was identified in 10 of the 347 controls (1.4%) and in 7 of 454 MS patients (0.8%; Table 1). No homozygous individuals were found either in the control or in the patient group. Genetic association between the *PTPRC* 77C \rightarrow G transition and MS susceptibility was excluded in the MS cohort. In addition, subgrouping patients according to differences in the clinical course of MS or according to *HLA-DRB1*15* status did not yield significant differences.

Conclusions: The 77C \rightarrow G transition in exon 4 of the *PTPRC* gene may contribute to MS susceptibility only in very few families, if at all, but it is not relevant for the majority of MS cases, including virtually all German patients.

Background

Multiple sclerosis (MS) is an autoimmune disease affect-

ing the central nervous system by demyelination. Both environmental and genetic components contribute to the

Table I: Allele frequency of the 77C \rightarrow G transition in PTPRC. Homozygosity was not observed.

| | C allele (%) | G allele (%) |
|-----------------------------------|--------------|--------------|
| Controls (N = 347) | 684 (98.6) | 10 (1.4) |
| MS patients (N = 454) | 901 (99.2) | 7 (0.8)* |
| RRMS (N = 219) | 435 (99.3) | 3 (0.7) |
| SPMS (N = 92) | 182 (98.9) | 2 (1.1) |
| PPMS $(N = 70)$ | 139 (99.3) | I (0.7) |
| HLA-DRB1*15 MS patients (N = 161) | 320 (99.4) | 2 (0.6) |

 * p = 0.2. RRMS = relapsing remitting course of MS; SPMS = secondary progressive course of MS; PPMS = primary progressive course of MS

development of MS. An efficient strategy to elucidate the genetic background of MS is to analyse single nucleotide polymorphisms (SNPs) in respective candidate genes. Since contradictory results have been reported recently [1-3], we analysed the 77C \rightarrow G transition in exon 4 of the protein-tyrosine phosphatase receptor-type C (PTPRC also known as CD45) [4]. PTPRC is localized on chromosome 1q31-q32 and consists of 35 exons. The gene encodes a 180-220 kDa glycoprotein expressed on leukocytes and hematopoietic progenitors [4]. This receptor is involved in T and B cell activation and in signal transduction by regulating protein-tyrosine kinases. Because of this involvement in immunological reactions, the gene is a candidate for MS predisposition. The protein exists in multiple isoforms, depending on alternative splicing of exons 4 (CD45RA), 5 (CD45RB) and 6 (CD45RC) (CD45RO, exon 4–6 spliced out). The 77C \rightarrow G transition does not change the amino acid sequence, but it is probably part of a motif necessary for splicing of CD45RA. The expression of CD45RA is increased in 77C/G heterozygous individuals [1].

Methods

Peripheral blood samples from more than 400 healthy blood donors were obtained with their informed consent, and provided by the department of transplantation and immunology of the University Hospital Eppendorf (Hamburg, Germany). The mean age of the healthy donors was 39.3 ± 11.47 years. The male/female ratio was 1.4 ± 0.5 . More than 800 unrelated MS patients attending the Department of Neurology, University clinics of Bochum and Göttingen (Germany) participated in this study. The male/female ratio was 1.7 ± 0.47 , the mean age at MS onset was 29.9 ± 9.63 years. 55.3% of all clinically diagnosed MS patients exhibited a relapsing/remitting course of MS (RRMS), 25.8% developed secondary progressive MS (SPMS) and 18.9% were characterized by primary progressive MS (PPMS). The mean EDSS (expanded disability status scale) of all MS patients was 4.2 ± 2.31 , of PPMS 5.8 \pm 1.91 and of RRMS+SPMS 3.9 \pm 2.25 [5].

Polymerase chain reaction (PCR) of exon 4 of PTPRC was carried out in a final volume of 12.5 µl with 50 ng of DNA, 200 µM dNTP, 1 U Taq Polymerase and PTPRC exon 4 specific primers (forward, 5'-ATTTATTTGTCCTTCTC-CCA-3' and reverse, 5'-GTTAACAACTTTTGTGTGCCAAC-3'). PCR cycling started with initial denaturation for 5 minutes at 94°C. The annealing temperature of the first cycle was 61°C, second cycle 58°C and remaining 26 cycles 55°C. The annealing time was 1 minute. Extension was performed at 72°C for 1 minute (final extension 5 minutes). PCR products were digested using endonuclease MspI according to the manufacturer's recommendation. Digested DNA was electrophoresed on 1.5% agarose gels. The restriction fragments of the wildtype C allele were 198 and 62 bp in length. The G allele harbors an additional restriction site so that the 198 bp fragment is digested into 114 and 84 bp (Figure 1).

Results

The rare G allele was present in 10 of the 347 controls (1.4%) and in 7 of 454 MS patients (0.8%; Table 1). There were no homozygous individuals either in the control or patient groups. Genetic association between the *PTPRC* 77C \rightarrow G transition and MS susceptibility was excluded in the MS cohort. In addition, subgrouping patients according to differences in the clinical course of MS (RRMS, SPMS, PPMS) or according to *HLA-DRB1*15* status did not yield significant differences.

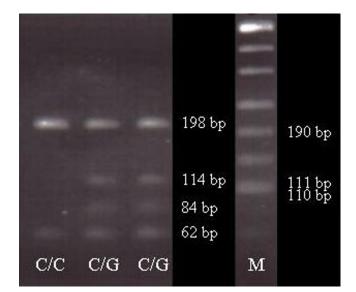


Figure I

Restriction fragment length polymorphism (RFLP) analysis of PTPRC exon 4 77C \rightarrow G; M=pUC19 DNA marker (501, 489, 404, 331, 242, 190, 147, 111, 110, 67 bp)

Discussion

This result contradicts the findings of Jacobsen *et al.*[1] who demonstrated an association of the mutation with MS in 3 families. Barcellos *et al.*[2] and Vorechovsky *et al.*[3] excluded MS association as well. However, as yet unknown polymorphisms in the *PTPRC* gene may contribute to MS susceptibility.

Conclusions

In conclusion, the *PTPRC* exon 4 77C \rightarrow G transition seems to contribute to MS susceptibility only in a few families, if at all, but it is not relevant for the majority of MS cases, also not in virtually all German patients.

Competing interests

None declared.

Authors' contributions

BM carried out the molecular analyses, ES, MH, and SS examined the MS patients, and JE participated in the study design and the coordination.

All authors have read and approved the final manuscript.

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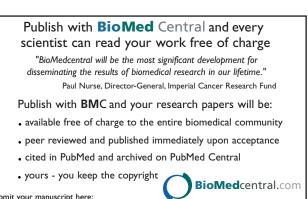
References

- Jacobsen M, Schweer D, Ziegler A, Gaber R, Schock S, Schwinzer R, Wonigeit K, Lindert R-B, Kantarci O, Schaefer-Klein J, Schipper HI, Oertel WH, Heidenreich F, Weinshenker BG, Sommer N, Hemmer B: A point mutation in PTPRC is associated with the development of multiple sclerosis. Nat Genet 2000, 26:495-499
- Barcellos LF, Caillier S, Dragone L, Elder M, Vittinghoff E, Bucher P, Lincoln RR, Pericak-Vance M, Haines JL, Weiss A, Hauser SL, Oksenberg JR: PTPRC (CD45) is not associated with the development of multiple sclerosis in U.S. patients. Nat Genet 2001, 29:2323-24
- Vorechovsky I, Kralovicova J, Tchilian E, Masterman T, Zhang Z, Ferry B, Misbah S, Chapel H, Webster D, Hellgren D, Anvret M, Hillert J, Hammarstrom L, Beverley PC: Does 77C→G in PTPRC modify autoimmune disorders linked to the major histocompatibility locus? Nat Genet 2001, 29:22-23
- Thude H, Hundrieser J, Wonigeit K, Schwinzer R: A point mutation in the human CD45 gene associated with defective splicing of exon A. Eur J Immunol 1995, 25:2101-2106
- Miterski B, Böhringer S, Klein W, Sindern E, Haupts M, Schimrigk S, Epplen JT: Inhibitors in the NFxB cascade comprise prime candidate genes predisposing to multiple sclerosis, especially in selected combinations. Genes and Immunity 2002

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